

Citation for published version:

Lowe, JD, Sanderson, HS, Cooke, AE, Ostovar, M, Tsisanova, E, Withey, SL, Chavkin, C, Husbands, SM, Kelly, E, Henderson, G & Bailey, CP 2015, 'Role of G Protein-Coupled Receptor Kinases 2 and 3 in μ -Opioid Receptor Desensitization and Internalization', *Molecular Pharmacology*, vol. 88, no. 2, pp. 347-356.
<https://doi.org/10.1124/mol.115.098293>

DOI:

[10.1124/mol.115.098293](https://doi.org/10.1124/mol.115.098293)

Publication date:

2015

Document Version

Peer reviewed version

[Link to publication](#)

University of Bath

Alternative formats

If you require this document in an alternative format, please contact:
openaccess@bath.ac.uk

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

**Role of G protein-coupled receptor kinases 2 and 3 in μ -opioid receptor
desensitization and internalization**

by

**Janet D. Lowe, Helen S. Sanderson, Alexandra E. Cooke, Mehrnoosh Ostovar, Elena
Tsisanova, Sarah L. Withey, Charles Chavkin, Stephen M. Husbands, Eamonn Kelly,
Graeme Henderson and Chris P. Bailey**

School of Physiology and Pharmacology, University of Bristol, Bristol, United Kingdom
(J.D.L., H.S.S., A.E.C., E.T., S. L. W., E.K., G.H.); Department of Pharmacology, University
of Washington School of Medicine, Seattle, WA, USA (C.C.); Department of Pharmacy and
Pharmacology, University of Bath, Bath, United Kingdom (M.O., S.H., C.P.B.)

Running Title: Role of GRKs in MOPr desensitization

Author for correspondence:

Dr C.P. Bailey

Department of Pharmacy and Pharmacology, University of Bath, Claverton Down, Bath, BA2
7AY, United Kingdom.

Tel: 44-1225-384957

Fax: 44-1225-386114

Email: c.p.bailey@bath.ac.uk

Number of

Text pages	38
------------	----

Tables	0
--------	---

Figures	5
---------	---

References	44
------------	----

Supplementary figures	2
-----------------------	---

Supplementary tables	1
----------------------	---

Number of words in

Abstract	212
----------	-----

Introduction	699
--------------	-----

Discussion	1424
------------	------

Non standard abbreviations

α_2 AR, α_2 adrenoceptor;

aCSF, artificial cerebrospinal fluid;

CaMKII, calcium/calmodulin-dependent protein kinase;

Cmpd101, Takeda compound 101;

CNQX, 6-cyano-7-nitroquinoxaline-2,3-dione;

DAMGO, D-Ala², *N*-MePhe⁴, Gly-ol]-enkephalin;

DMEM, Dulbecco's modified Eagles's medium;

DNM, dominant negative mutant;

ECL, enhanced chemiluminescence;

ERK, extracellular-signal-regulated kinase;

GIRK channel, G protein-activated potassium channel;

GRK, G protein-coupled receptor kinase;

HA, haemagglutinin;

HEK 293 cells, human embryonic kidney 293 cells;

HRP, horseradish peroxidase;

JNK, c-Jun N-terminal kinase;

KO, knock out;

LC, locus coeruleus;

MOPr, μ opioid receptor;

NA, noradrenaline (norepinephrine);

PDB, pull down buffer;

PKA, protein kinase A;

PKC, protein kinase C;

PMSF, phenylmethanesulfonylfluoride;

PRK2, protein kinase C related protein kinase;

PVDF, polyvinylidene fluoride;

ROCK2, Rho-associated protein kinase 2;

MOL #98293

SDS, sodium dodecyl sulphate;

SGK1, serum and glucocorticoid regulated kinase (SGK1);

SST, somatostatin;

SST2R, somatostatin 2 receptor;

WGA, wheat germ agglutinin;

WT, wild type

Abstract

There is ongoing debate about the role of G protein receptor kinases (GRKs) in agonist-induced desensitization of the μ opioid receptor (MOPr) in brain neurons (see Williams et al., 2013, Pharmacol Rev 15;223-254). In the present paper we have used a novel, membrane permeable, small molecule inhibitor of GRK2 and GRK3, Takeda Compound 101 (Cmpd101), to study the involvement of GRK2/3 in acute agonist-induced MOPr desensitization. We observed that Cmpd101 inhibits the desensitization of the G protein activated potassium current (GIRK) evoked by receptor- saturating concentrations of methionine enkephalin (Met Enk), D-Ala²,N-MePhe⁴,Gly-ol]-enkephalin (DAMGO), endomorphin-2 and morphine in rat and mouse locus coeruleus (LC) neurons. In LC neurons from GRK3 knockout mice Met Enk-induced desensitization was unaffected implying a role for GRK2 in MOPr desensitization. Quantitative analysis of the loss of functional MOPrs following acute agonist exposure revealed that Cmpd101 only partially reversed MOPr desensitization. Inhibition of ERK1/2, PKC, JNK or GRK5 did not inhibit the Cmpd101-insensitive component of desensitization. In HEK 293 cells Cmpd101 produced almost complete inhibition of DAMGO-induced MOPr phosphorylation at Ser³⁷⁵, arrestin translocation and MOPr internalization. Our data demonstrate a role for GRK2 (and potentially also GRK3) in agonist-induced MOPr desensitization in the LC, but leave open the possibility that another, as yet unidentified, mechanism of desensitization also exists.

Introduction

The canonical pathway for homologous G protein-coupled receptor (GPCR) desensitization involves agonist activation of the receptor, subsequent G protein activation, G protein-coupled receptor kinase (GRK) translocation to the plasma membrane, phosphorylation of serine and threonine residues primarily on the C-terminal tail of the receptor by GRKs and arrestin binding to the phosphorylated receptor (Kelly et al., 2008). While binding of arrestins to the receptor prevents further G protein activation (i.e. desensitizes the receptor-G protein-coupled effector activation pathway) it may also lead to arrestin-mediated signalling and receptor internalization, recycling and downregulation (Shenoy & Lefkowitz, 2011).

In LC neurons taken from relatively mature animals DAMGO, a high efficacy MOPr agonist, induces substantial homologous MOPr desensitization (Llorente et al., 2012). However, despite considerable effort there still remains controversy and confusion around the role of GRKs in the rapid desensitization of neuronal μ -opioid receptors (MOPrs) (for extensive review, see Williams et al., 2013). Evidence in favour of a role of GRKs in neuronal MOPr desensitization by agonists with high intrinsic efficacy is threefold. First DAMGO-induced MOPr desensitization in rostral ventromedial medulla neurons was shown to be inhibited by intracellular perfusion of a GRK2 inhibitory peptide (Li and Wang, 2001). Second, fentanyl-induced MOPr desensitization in the hippocampal dentate gyrus was absent in brain slices from GRK3 knockout (KO) mice (Terman et al., 2004). Third, overexpression of a GRK2 dominant negative mutant (DNM) in rat LC neurons reduced DAMGO-induced MOPr desensitization (Bailey et al., 2009a). On the other hand two studies have failed to provide evidence for any involvement of GRKs in MOPr desensitization. Neither heparin, a low affinity GRK inhibitor, nor staurosporine, a non-selective kinase inhibitor, had any effect on Met Enk-induced MOPr desensitization in rat LC neurons (Arttamangkul et al., 2012). Also, using the GRK2as5 transgenic mouse in which GRK2 has been mutated to allow chemical inhibition it was observed that exposure to the inhibitor did not reduce Met Enk-induced

desensitization in LC neurons (Quillinan et al., 2011). However, this transgenic knock-in/chemical inhibition approach would only inhibit GRK2 and not affect GRK3 or other GRKs.

Further complicating the issue, Dang et al., (2009) reported that in rat LC neurons both GRKs and ERKs were involved in MOPr desensitization; inhibition of GRK2/3 by intracellular dialysis of inhibitory peptides or ERK1/2 with a small molecule ERK1/2 activation inhibitor alone did not reduce Met Enk-induced desensitization but with concomitant inhibition of both types of kinase desensitization was inhibited.

In LC neurons *in vitro* morphine, an agonist with lower intrinsic efficacy (McPherson et al., 2010), induced much less MOPr desensitization than high efficacy opioid agonists (Alvarez et al., 2002; Bailey et al., 2003). The level of morphine-induced desensitization could be enhanced by concomitant activation of PKC either indirectly by stimulation of Gq-coupled M₃ muscarinic receptors or directly with a phorbol ester (Bailey et al., 2004). Expression of the GRK2 DNM did not prevent PKC-mediated MOPr desensitization on morphine application (Bailey et al., 2009a). In contrast, in hippocampal dentate gyrus GRK3 knockout reduced morphine-induced desensitization (Terman et al., 2004).

Until very recently there have been no small molecule selective, membrane permeable inhibitors of GRKs. In this paper we have examined the effects of a novel small molecule inhibitor of GRK2 and GRK3, Takeda Compound 101 (Cmpd101) (Ikeda et al., 2007; Thal et al., 2011), as well as GRK3 knockout and signalling kinase inhibitors on the rapid desensitization of the MOPr-activated inwardly rectifying potassium current (GIRK) in LC neurons. We have sought to address several questions relating to the role of GRKs in MOPr desensitization. First, are GRK2 and GRK3 involved in the acute MOPr desensitization induced by high intrinsic efficacy agonists, arrestin-biased agonists and lower intrinsic efficacy agonists? Second, do both GRK2 and GRK3 play a role in MOPr desensitization?

Third, are multiple kinases such as GRKs and ERKs involved in acute MOPr desensitization?

Our results suggest that GRK2, and potentially GRK3, but not ERK1/2 or JNK are involved in MOPr desensitization by all of the MOPr agonists studied. However, even at concentrations of Cmpd101 that almost completely inhibited MOPr phosphorylation at Ser³⁷⁵ and MOPr internalization in HEK 293 cells the inhibition of agonist-induced loss of functional MOPr in LC neurons was only of the order of 34%.

Materials and Methods

Brain slice preparation

Male Wistar Rats (5 - 8 weeks old) were killed by cervical dislocation and male mice (wild type and GRK3 knockout mice backcrossed onto C57Bl/6 background, 5 - 8 weeks old) were decapitated under isoflurane-induced anaesthesia. Brains were removed and submerged in ice-cold cutting solution containing (in mM): 20 NaCl, 2.5 KCl, 0.5 CaCl₂, 7 MgCl₂, 1.25 NaH₂PO₄, 85 sucrose, 25 D-glucose and 60 NaHCO₃, saturated with 95% O₂/5% CO₂. Horizontal brain slices (230 µm thick) containing the LC were prepared using a vibratome (VT1000S, Leica Biosystems, Milton Keynes, UK). Slices were then transferred to artificial cerebrospinal fluid (aCSF) composed of (in mM): 126 NaCl, 2.5 KCl, 1.2 MgCl₂, 2.4 CaCl₂, 1.2 NaH₂PO₄, 11.1 D-glucose, 21.4 NaHCO₃, and 0.1 ascorbic acid, saturated with 95% O₂/5% CO₂ at 33 °C, and were left to equilibrate for at least 1 h prior to recording. All experiments were performed in accordance with the UK Animals (Scientific Procedures) Act 1986, the European Communities Council Directive 1986 (86/609/EEC) and the University of Bristol and University of Washington ethical review documents as appropriate.

Whole-cell path-clamp recording

Slices were submerged in a slice chamber (Warner Instruments, Hamden CT, USA) mounted on a microscope stage (BX51WI, Olympus, Southend-on-Sea, UK) and superfused (2.5 - 3 ml/min) with aCSF at 33 °C. LC neurons were visualized using Nomarski optics using infrared light. Whole-cell patch clamp recordings were made using electrodes (3-6 MΩ) filled with (in mM): 115 K-methylsulfonate, 10 NaCl, 2 MgCl₂, 10 HEPES, 6 EGTA, 2 MgATP, and 0.5 Na₂GTP (pH 7.25, osmolarity 270 mOsm). Recordings of whole-cell currents were filtered at 1 kHz using an Axopatch 200B amplifier (Molecular Devices, Sunnyvale CA, USA) and analysed off-line using WinEDR and WinWCP (University of Strathclyde, Glasgow, UK), and pClamp (Molecular Devices, Sunnyvale CA, USA).

LC neurons were voltage clamped at -60 mV with correction for a -12 mV liquid junction potential. Activation of MOPr receptors and α₂ adrenoceptors (α₂ AR) evoked inwardly rectifying potassium channel (GIRK) currents. All drugs were applied in the

superfusing solution in known concentrations. In those experiments in which α_2 AR responses were studied, noradrenaline (NA) was applied in the presence of prazosin (1 μ M) and cocaine (3 μ M). Because GIRK currents were much smaller in mouse LC, responses in mice were measured in the presence of the glutamate receptor antagonist 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX, 10 μ M) and the GABA_A receptor antagonist bicuculline (10 μ M) to inhibit spontaneous excitatory and inhibitory postsynaptic currents.

Cell culture

Human embryonic kidney 293 cells (HEK 293) cells stably over expressing haemagglutinin-tagged rat MOPr (HA-MOPr) were cultured at 37 °C in 5% CO₂ in Dulbecco's modified Eagles's medium (DMEM) supplemented with 10 % fetal bovine serum, 10 U/ml penicillin, 10 mg/ml streptomycin (Invitrogen, Life Technologies, Paisley UK) and 250 μ g/ml G418 (Merck, Nottingham, UK). Cells were seeded onto 60 mm dishes and grown to 90 % confluence then subjected to serum starvation for 24 h.

WGA-enrichment of HA-MOPr

Glycosylated HA-MOPr was enriched from cells that were lysed in pull down buffer (PDB) composed of (in mM) 20 HEPES [pH 7.4], 150 NaCl, 1 Na₃VO₄, 10 β -glycerophosphate, 50 NaF, 1 PMSF and 5 EDTA plus 1% Triton X-100, 10% glycerol, 0.1% SDS, 2 μ g/ml leupeptin, 2 μ g/ml aprotinin and 1 μ g/ml pepstatin A. The protein concentration of clarified cell lysate was adjusted to 1.5 mg/ml and an input sample removed for Western blot analysis. 2.25 mg of cell lysate was then incubated with 15 μ l sedimented wheat germ agglutinin (WGA)-agarose beads (L1394, Sigma Aldrich, Dorset UK), prewashed in PDB and incubated overnight at 4 °C. Beads were then washed three times in PDB for 5 min at 4 °C. WGA-binding proteins were eluted with the addition of SDS-sample buffer and boiling at 95 °C for 4 min.

Western blotting

Eluted WGA-binding proteins and input sample were supplemented with β -mercaptoethanol for gel loading. Samples were subject to SDS polyacrylamide gel electrophoresis and proteins transferred onto polyvinylidene fluoride (PVDF) membranes. For detection of

agonist-induced receptor phosphorylation antibodies targeting pSer³⁷⁵ on MOPr were used (dilution 1:1000, Cell Signalling Technology, Danvers, MA) and total receptor levels detected with anti-HA antibody (MMS-101R-200, 1:1000, Cambridge Bioscience Limited, Cambridge UK). For input samples, anti-pERK1/2, anti-ERK1/2 and anti-pElk-1 (#9101, #9107, #9181 each 1:1000, Cell Signalling Technology) and anti-tubulin antibodies were used (T6074, 1:10000, Sigma Aldrich). Following incubation with the species appropriate horseradish peroxidase (HRP)-conjugated secondary antibody (NA934V, donkey anti-rabbit, dilution 1:7500; NA931V, sheep anti-mouse, 1:10000, GE Healthcare, Buckinghamshire UK), bands were visualized by enhanced chemiluminescence (ECL) with SuperSignal West Dura Chemiluminescent Substrate (Thermo Fisher Scientific, Hemel Hempstead, UK). Densitometry of bands was undertaken using ImageJ (NIH, USA), duplicate values were taken for each sample and then averaged. pSer³⁷⁵ levels were normalized against corresponding total HA-MOPr levels and pERK1/2 against total ERK1/2 levels determined in the same experiment.

Internalization Assays

DAMGO-induced internalization of HA-MOPr was assessed by ELISA as previously described (Johnson et al., 2006) and by confocal microscopy imaging (Mundell et al., 2006). HEK 293 cells stably expressing HA-MOPr were prelabelled with primary antibody for 1 h at 4 °C before incubation with Cmpd101 (3 or 30 µM) for 30 min at 37 °C. Cells were then stimulated with DAMGO (10 µM) at 37 °C to induce internalization. In the ELISA changes in surface receptor expression were subsequently determined by normalizing data from each treatment group to corresponding control surface receptor levels determined from cells not exposed to DAMGO in the same experiment. For confocal imaging cells were imaged using a Leica SP5-AOBS confocal laser scanning microscope attached to a Leica DM16000 inverted epifluorescence microscope with a pLApoBL 63 x oil immersion objective.

Arrestin Translocation Assay

For the arrestin-3 recruitment assay, PathHunter™ cells and accompanying assay kit were obtained from DiscoverX (Birmingham, UK). These cells are U2OS (human osteosarcoma)

cells stably expressing tagged human MOPr and tagged arrestin-3 such that recruitment of arrestin-3 to the receptor leads to reconstitution of β -galactosidase activity which is measured by luminescence in a plate reader (McPherson et al., 2010). The experiments were undertaken exactly as described in the manufacturer's instructions, with the agonist DAMGO being added to the cells for 90 min, and where applicable Cmpd101 being added alone or 30 min before DAMGO.

Kinase Screen

The express kinase screen was performed at the Medical Research Council International Centre for Kinase Profiling (Dundee, UK). The screen was comprised of 52 human enzymes providing a representative sampling of the kinome, with the exception of lipid kinases. The method used was a radioactive filter binding assay using 32 ATP which provides a direct measure of kinase activity (Hastie et al., 2006; Bain et al., 2007). Cmpd101 was studied at a concentration of 1 μ M. Results are given as the percentage of kinase activity in the absence of the inhibitor. The ATP concentrations used were at or below the calculated K_m for ATP for each individual kinase. The screen was performed in duplicate and results are given as a mean value with standard deviation.

Quantification of DAMGO-induced loss of functional MOPr

Following desensitization of MOPr by bath-application of DAMGO, DAMGO was replaced by morphine. Using the operational model of agonism (Black et al., 1985), transducer ratio (τ) values for morphine were calculated (as in Bailey et al., 2009b). Percentage loss of receptor responsiveness (f) was then calculated using the equation $f = 100 \times (1 - \tau_2/\tau_1)$ where τ_1 is the value in control slices and τ_2 is the value after DAMGO-induced desensitization (Lohse et al., 1990).

Statistics

Data are presented as means \pm standard errors and were analysed by unpaired two-tailed Student's t-tests, one-way or two-way ANOVA with Bonferroni post-hoc test as appropriate using Prism5 (Graphpad Software Inc., San Diego CA, USA). Differences were assumed to be significant at $P < 0.05$.

Drugs and compounds

3-[[[4-Methyl-5-(4-pyridyl)-4H-1,2,4-triazole-3-yl] methyl] amino]-N-[2-(trifluoromethyl) benzyl] benzamidehydrochloride (Cmpd101 hydrochloride) was synthesized at the University of Bath (Bath, UK) in 9 steps from isonicotinic acid hydrazide (Ikeda et al., 2007). Analysis of the final compound (as the free base) provided data consistent with the structure with purity >99% (by ^{13}C nmr): ^1H NMR (400 MHz, CDCl_3) δ 8.71 (dd, $J = 1.6$ Hz, $J = 4.4$ Hz, 2H); 7.63-7.45 (m, 5H); 7.34 (t, $J = 7.6$ Hz, 1H); 7.25-7.24 (m, 1H); 7.18 (t, $J = 7.6$ Hz, 1H); 7.06 (d, $J = 6.8$ Hz, 1H); 6.91-6.85 (m, 2H); 4.76 (d, $J = 6.00$ Hz, 2H), 4.50 (s, 2H); 3.71 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 167.6, 153.8, 153.2, 150.3, 147.5, 136.6, 135.2, 134.4, 132.3, 130.2, 129.5, 128.1, 127.8, 127.5, 126.0, 125.9, 125.8, 125.7, 123.1, 122.5, 116.6, 116.3, 111.9, 76.7, 40.5, 39.3, 31.6, ESIMS: calc. for $\text{C}_{24}\text{H}_{21}\text{F}_3\text{N}_6\text{NaO}$, 489.1627; found 489.1701.

D-Ala²,N-MePhe⁴,Gly-ol]-enkephalin (DAMGO) and methionine enkephalin (Met Enk), (Bachem, Bubendorf, Switzerland); morphine hydrochloride (Macfarlane-Smith, Edinburgh UK); cocaine hydrochloride (Sigma Aldrich); endomorphin-2, bicuculline, 6-cyano-7-nitroquinoxaline-2,3-dione disodium (CNQX), naloxone hydrochloride, noradrenaline bitartrate, prazosin hydrochloride, Y-27632, amlexanox (Ascent Scientific / Abcam Biochemicals, Cambridge, UK); GF109203X, GSK 650384, PD 98059 (Tocris Bioscience, Bristol, UK). All other reagents were from Sigma Aldrich.

RESULTS

Role of GRK2 and GRK3 in acute MOPr desensitization induced by Met Enk or DAMGO.

In rat LC neurons, activation of MOPrs by receptor-saturating concentrations of Met Enk (30 μ M), DAMGO (10 μ M) or endomorphin-2 (10 μ M) elicited outward current through GIRK channels that rapidly desensitized in the presence of the drug, with only about 50% of the initial current remaining after 10 min exposure (Fig. 1). To examine the role of GRK2 and GRK3 in the Met Enk-induced desensitization, LC slices were pre-treated with the inhibitor, Cmpd101, at concentrations previously shown to inhibit desensitization of β_2 adrenoceptors in HEK 293 cells (Ikeda et al., 2007). Cmpd101 concentration-dependently reduced the amount of Met Enk-induced desensitization (Fig. 1A-C) such that in the presence of Cmpd101 (30 μ M) the desensitization induced by Met Enk (30 μ M) was reduced to only 15%. We also observed that Cmpd101 reduced the amount of desensitization induced by DAMGO (10 μ M, Fig. 1D-F) and endomorphin-2 (10 μ M, Fig. 1G-I).

To determine whether there was any additional role for ERK1/2, PKC or JNK in MOPr desensitization we exposed cells to Cmpd101 and PD98059 (10 μ M) to inhibit ERK1/2 (Met Enk as agonist, Fig. 1C), SP600125 (30 μ M) to inhibit JNK (Met Enk as agonist, Fig. 1C), or GF109203X (1 μ M) to inhibit PKC (DAMGO as agonist, Fig. 1F). We did not observe any further reduction in desensitization in the presence of the ERK1/2, PKC, or JNK inhibitors. Neither the ERK1/2 nor JNK inhibitor applied on its own in the absence of Cmpd 101 affected Met Enk-induced desensitization (Control: $51 \pm 6\%$, $n=5$; + 10 μ M PD98059: $43 \pm 5\%$, $n=5$; + 30 μ M SP600125: $50 \pm 4\%$, $n=5$). Amlexanox has recently been identified as a GRK5 inhibitor (Homan et al., 2014b). When cells were exposed to Cmpd101 and amlexanox (50 μ M) there was no further reduction in desensitization compared with Cmpd101 alone (Fig. 1C). Amlexanox applied on its own in the absence of Cmpd101 did not affect Met Enk-induced desensitization (% Met Enk-induced desensitization in 50 μ M amlexanox alone: $51 \pm 3\%$, $n=5$).

The inhibition of MOPr desensitization that we observed with Cmpd101 does not appear to be due to it acting as a MOPr antagonist to reduce the number of receptors being activated. When a submaximal concentration of Met Enk (600nM) was applied before and following exposure of cells to Cmpd101 (3 μ M for 15 min), the Met Enk response in the presence of Cmpd101 was not different from the initial response before Cmpd101 ($94 \pm 13\%$ of initial response, $n=4$, one-sample t-test vs. 100%, $p>0.05$).

Role of GRK2 and GRK3 in morphine-induced MOPr desensitization

Although morphine produces much less acute MOPr desensitization in LC neurons compared to either Met Enk or DAMGO (Bailey et al., 2004; 2009a), the morphine response did desensitize by about 25% over a 15 min application (Fig. 1J-L). Pre-treatment with Cmpd101 (30 μ M) significantly reduced the morphine-induced desensitization. This suggests that GRK2 and GRK3 are not only involved in desensitization to higher efficacy MOPr agonists such as Met Enk and DAMGO, but are also involved in desensitization to the partial agonist morphine in the absence of PKC activation.

MOPr desensitization in LC neurons from GRK3 knockout mice

Cmpd101 was previously shown to be almost equally effective at inhibiting GRK2 and GRK3 (Thal et al., 2011). GRK3 KO has been reported to abolish the desensitization induced by fentanyl and morphine in rat hippocampal dentate gyrus neurons (Terman et al., 2004). To determine whether GRK3 was the sole isoform required for MOPr desensitization in LC neurons, we measured Met Enk-induced desensitization in LC neurons prepared from GRK3 KO mice. The amplitude of the Met Enk-induced current was similar in neurons from both GRK3 KO and wild type (WT) mice (WT: 72 ± 10 pA, $n=6$; GRK KO: 58 ± 11 pA, $n=6$, $p>0.05$). Met Enk-induced desensitization measured after 10 min of Met Enk application was also not significantly different in GRK3 KO neurons compared to WT controls (Fig. 2A-B). To ensure that there was not a species-specific difference in the mechanism of desensitization,

we also examined the ability of Cmpd101 to inhibit Met Enk-induced desensitization in mouse LC neurons since the previous experiments with Cmpd101 were performed in rat LC neurons. In LC neurons from C57Bl/6J WT mice, Met Enk produced a similar amount of desensitization (45%) to that observed in rat neurons after a 10 min application. Cmpd101 inhibited the Met Enk-induced desensitization in WT mouse LC neurons, being maximally effective at the lower (3 μ M) concentration (Fig. 2C-D).

These results suggest first that GRK2 plays a role in Met Enk-induced desensitization in LC neurons and second, either that GRK3 plays no role in Met Enk-induced desensitization in LC neurons or that GRK3 is not solely responsible. However, as Cmpd101 is a mixed GRK2/3 inhibitor, we cannot exclude the possibility that both GRK2 and GRK3 are involved and that removal of only one (in our current experiments GRK3) is insufficient to inhibit desensitization.

Loss of functional MOPrs underlying acute MOPr desensitization

There is a significant MOPr reserve in LC neurons, that needs to be removed (i.e. desensitized) before any decrease in the response to a supramaximal concentration of a full agonist such as Met Enk or DAMGO can be detected and therefore the true proportion of receptors that have been desensitized is hard to assess from responses such as those shown in Fig. 1 (Connor et al., 2004). To determine the actual loss of functional MOPrs following desensitization we used a previously developed protocol that measures the decrease induced by desensitization of the maximum response to the partial agonist morphine (Bailey et al., 2009b). For a partial agonist, the maximum response occurs when all the receptors are occupied and therefore any decrease in maximum response is directly related to a loss of receptor function. To be able to compare results from different neurons, the morphine response in each was normalized to the response to noradrenaline (NA, 100 μ M), which activates α_2 ARs that couple to the same pool of GIRKs in LC neurons (North and Williams, 1985).

In control LC neurons the maximum response evoked by morphine was 85% of the maximum response to NA (100 μ M) (Fig. 3A). To examine the reduction in morphine response caused by DAMGO-induced MOPr desensitization, neurons were first treated with DAMGO (10 μ M) for 12 min to induce desensitization, then the maximum response to morphine (30 μ M) was determined at plateau (6 mins after DAMGO was replaced by morphine; Fig. 3B & C). In neurons that were first treated with DAMGO (10 μ M) for 12 min to induce desensitization the maximum response to morphine was reduced by over 80% (Fig. 3B). Using the operational model of agonism (Black et al., 1985) to estimate the actual loss of functional MOPrs induced by DAMGO (for details see Bailey et al., 2009b and Methods section) we calculate that the DAMGO treatment induced a 93% loss of functional MOPrs. When we repeated the experiment but this time in the presence of Cmpd101 (3 or 30 μ M) the desensitization induced by DAMGO was markedly reduced as was the reduction in the maximum response to morphine (Fig. 3C & D). In the presence of 30 μ M Cmpd101 the desensitization induced by DAMGO (10 μ M) reduced the maximum response to morphine by only 44% which equates to a 59% loss of functional MOPrs. Thus whilst Cmpd101 reduced the DAMGO-induced loss of functional receptors from 93 to 59% this reduction was sufficient to produce a marked reduction in the desensitization observed during application of a receptor-saturating concentration of agonist (Fig. 1).

Overall our findings with Cmpd101 suggest that GRK2 and possibly also GRK3 are involved in both Met Enk- and DAMGO-induced desensitization of MOPr, but it is clear that there is a component of desensitization that still remains in the presence of the drug either because of incomplete inhibition of GRK2 and GRK3 by Cmpd101 or because an additional Cmpd101-resistant mechanism of desensitization exists.

MOPr phosphorylation, internalization and arrestin translocation

To assess the ability of Cmpd101 to inhibit GRK in intact cells we next examined agonist-induced MOPr phosphorylation, arrestin translocation and MOPr internalization. Because these processes are more difficult to assess in neurons in a brain slice, we instead examined them in cell lines stably expressing MOPrs. Phosphorylation of HA-tagged MOPr was measured by Western blotting using the commercially available antibody that recognizes the phosphorylated residue Ser³⁷⁵ in the MOPrC-terminal tail. We have previously demonstrated that *in vitro* GRK2 directly phosphorylates Ser³⁷⁵ (Chen et al., 2012). Application of DAMGO (10 μ M) for 5 min produced a robust phosphorylation of Ser³⁷⁵ that was partially inhibited by pre-treatment of cells for 30 min with 3 μ M Cmpd101 and fully blocked by pre-treatment with 30 μ M Cmpd101 (Fig. 4A & B). Cmpd101 (30 μ M) also prevented phosphorylation of MOPr at Thr³⁷⁰, Thr³⁷⁶ and Thr³⁷⁹ residues also known to be phosphorylated by GRKs (Sanderson & Kelly, unpublished observations).

Translocation of arrestin3 to agonist-activated MOPr was assessed using the DiscoverX PathHunter^R assay. Application of DAMGO (10 μ M for 90min) led to a large recruitment of arrestin3 to MOPr which was dramatically reduced by in the presence of Cmpd101 (30 μ M) (Fig. 4C).

We assessed internalization of HA-tagged MOPrs by ELISA and confocal microscopy using an anti-HA antibody to label surface receptors. DAMGO (10 μ M) application induced MOPr internalization in a time-dependent manner and Cmpd101 markedly reduced DAMGO-induced MOPr internalization (Fig. 4D& E).

These data demonstrate that Cmpd101 was able to block DAMGO-induced MOPr phosphorylation on Ser³⁷⁵, arrestin recruitment, and MOPr internalization presumably by inhibiting GRK2 and GRK3. This contrasts somewhat with our data from LC neurons where although Cmpd101 could reduce agonist-induced desensitization the extent of the reduction was only about 34% when calculated as loss of functional receptors.

ERK1/2 phosphorylation

It has previously been reported that GRK2 and GRK3 inhibition alone was not sufficient to inhibit Met Enk-induced desensitization in mouse LC neurons but that a combination of GRK2, GRK3 and ERK1/2 inhibition was required i.e. there was overlap and redundancy between GRK and ERK1/2 in MOPr desensitization (Dang et al., 2009). To confirm that Cmpd101 did not inhibit ERK1/2 activity, we measured both the phosphorylation of ERK1/2, which is often used as a measure of its activity (Roskoski, 2012) and the phosphorylation of Elk-1 by ERK1/2 (Cruzalegui et al., 1999) in HEK 293 cells stably expressing HA-tagged MOPrs. Cmpd101 did not affect the DAMGO-induced (10 μ M, 5 min) increase in ERK1/2 phosphorylation (Fig. 5A & B). Moreover, at 30 μ M Cmpd101 on its own produced a small increase in basal ERK1/2 phosphorylation (Fig. 5A & B). Similarly, Cmpd101 (30 μ M) did not inhibit the DAMGO-induced (10 μ M, 5 min) increase in Elk-1 phosphorylation (Fig. 5C & D). The ERK1/2 activation inhibitor, PD98059, significantly decreased both DAMGO-induced ERK1/2 and Elk-1 phosphorylation (Fig. 5E).

Thus, the ability of Cmpd101 to decrease MOPr desensitization in the LC is unlikely to be due to an inhibition of ERK1/2 activity in combination with the inhibition of GRK activity.

Selectivity of kinase inhibition by Cmpd101

Cmpd101 has previously been shown to be highly selective for GRK2 and GRK3 over other GRK isoforms (Ikeda et al., 2007; Thal et al., 2011). We subjected Cmpd101 to a further kinase screen at the Medical Research Council International Centre for Kinase Profiling (Dundee, UK) to determine whether it inhibited the activity of other non-GRK kinases. As this screen was conducted on purified kinases *in vitro* we performed the screen using 1 μ M Cmpd101 (i.e. 30x the reported IC₅₀ for inhibition of GRK2 in *in vitro* purified enzyme assay conditions). Of the kinases that have previously been implicated in MOPr desensitization, Cmpd101 inhibited the activity of PKC, PKA, JNK, CaMKII and ERK1/2 *in vitro* by less than

20% at 1 μ M (Supplementary Data Table 1 and Supplementary Figure 1). Cmpd101 inhibited only 5 off-target kinases by more than 50%; these included protein kinase C related protein kinase (PRK2) and serum and glucocorticoid regulated kinase (SGK1) which were inhibited by 93% and 69% respectively. To test whether either of these kinases might be involved in DAMGO-induced desensitization, we pre-treated rat LC slices with established inhibitors of these kinases. GSK650394 (10 μ M) is an inhibitor of SGK1 and Y-27632 (50 μ M) is an inhibitor of both PRK2 and ROCK2 which Cmpd101 also inhibited by 47% in the kinase screen. LC neurons pre-treated with a combination of these two kinase inhibitors did not show any difference in the amount of desensitization induced by a 10 min application of DAMGO (30 μ M) compared to that observed in control neurons (Supplementary Figure 2). Therefore, these off target actions of Cmpd101 do not appear to play a role in the inhibition of MOPr desensitization by Cmpd101.

Discussion

We have studied the effects of Cmpd101, a novel membrane permeable, small molecule inhibitor of both GRK2 and GRK3 on agonist-induced MOPr desensitization in LC neurons. Cmpd101 has previously been reported to be a potent inhibitor of GRK2 and GRK3, with reported IC_{50} values of 35 nM (Ikeda et al., 2007) or 290 nM (Thal et al., 2011) under different *in vitro* purified enzyme assay conditions, but has no activity against GRK5 at concentrations up to 125 μ M (Thal et al., 2011). In intact cell assay conditions, higher concentrations are required, with Cmpd101 showing concentration-dependent inhibition of β_2 adrenoceptor desensitization in HEK 293 cells over the range 3 – 30 μ M (Ikeda et al., 2007).

In our preliminary kinase screen Cmpd101 showed high selectivity when tested across a broad range of kinases with only the AGC kinases, PRK2, MSK1 and SGK1, being inhibited by over 50% in the presence of 1 μ M Cmpd101. These kinases all sit in regions of the kinome (Manning et al., 2002) close to GRK2 and GRK3 (see Supplementary data Fig. 1), although PRK2 and SGK1 appear to play no role in MOPr desensitization (see Supplementary data Fig. 2). Other approaches previously taken to inhibit GRKs include the broad spectrum kinase inhibitor, staurosporine (Arttamangkul et al., 2012), the PKC/GRK inhibitor Ro32-0432 (Hull et al., 2010), ' β -ARK-1 inhibitor' (Iino et al., 2002; Hull et al., 2010) and the antidepressant drug paroxetine (Thal et al., 2012; Homan et al., 2014a). However, none of these approaches combine the cell-permeable, potency and selectivity profiles of Cmpd101. Moreover, Cmpd101 shows specificity between GRK2/3 and GRK5. Across a range of opioid agonists with different intrinsic efficacies for G protein activation (Met Enk, DAMGO and morphine) and arrestin bias (endomorphin 2) we observed that Cmpd101 reduced the desensitization of the opioid-activated GIRK current indicating the involvement of GRK2 and/or GRK3 in this desensitization. The effective concentrations of Cmpd101 in these whole cell assays was 100 – 1000 fold higher than the concentrations that inhibit GRK2 and GRK3 *in vitro*. This likely results from low permeability of the molecule across the

cell membrane but could also be due to other factors such as differences in the conformation or properties of the kinases in a cellular environment.

Our observation that MOPr desensitization in LC neurons was unaffected by GRK3 knockout indicates that in these neurons GRK3 is not solely responsible for MOPr desensitization. This observation is different from what has been reported previously in the hippocampal dentate gyrus where fentanyl-induced MOPr desensitization was absent in brain slices prepared from GRK3 KO mice (Terman et al., 2004). Suggesting that different mechanisms of MOPr desensitization may be present in different brain regions. This difference could result from differential expression of GRK isoforms in different neuronal populations. In situ hybridisation studies in rats have shown ubiquitous expression of GRK2, 3 and 6 throughout the brain (Erdtmann-Vourliotis et al, 2001), with expression of GRK2, 3, 5 and 6 in the LC but no studies have performed quantitative analysis of GRK isoform protein levels in different brain regions. Although GRK2 KO is embryonic lethal, selective inhibition of GRK2 in the GRK2as5 transgenic mouse has also been reported not to affect Met Enk-induced desensitization in LC neurons (Quillinan et al., 2011). Further, although overexpression of a GRK2 dominant negative mutant (DNM) reduced DAMGO-induced MOPr desensitization in rat LC neurons (Bailey et al., 2009a), this DNM construct is likely to also inhibit GRK3, based on homology of the GRK2 and GRK3 isoforms. Given that Cmpd101 is a potent inhibitor of both GRK2 and GRK3 our data suggest that in LC neurons there is an overlap and redundancy of GRK2 and GRK3 such that KO of only one (e.g. GRK3) is insufficient to inhibit desensitization.

Cmpd101 inhibited agonist-induced phosphorylation of MOPr at Ser³⁷⁵ and the subsequent internalization of the receptor in HEK 293 cells. Several groups have previously demonstrated that GRK2 and GRK3 rapidly phosphorylate MOPr. Purified GRK2 phosphorylates Ser³⁷⁵ in the C-terminal tail *in vitro* (Chen et al., 2012). Using phosphosite-specific antibodies DAMGO and other higher efficacy agonists have been observed to

phosphorylate several amino acid residues in the C-terminal tail of MOPr (Doll et al., 2011, 2012; Just et al., 2013). Phosphorylation at Ser³⁷⁵ appears to be the initiating event but thereafter flanking residues (Thr³⁷⁰, Thr³⁷⁶, and Thr³⁷⁹) are subsequently phosphorylated in a hierarchical manner. This higher-order phosphorylation has been shown to be mediated by GRK2 and GRK3 in that it is reduced by siRNA knockdown of these GRK isoforms. Furthermore, mutation of Ser³⁷⁵, Thr³⁷⁶ and Thr³⁷⁹ to Ala prevented DAMGO-induced MOPr internalization in medium spiny striatal neurons (Just et al., 2013). Morphine, a MOPr partial agonist in the LC, has previously been shown to produce much less acute MOPr desensitization in LC neurons (Alvarez et al., 2002; Bailey et al., 2003) than high efficacy MOPr agonists. It has also been shown to induce less phosphorylation of Ser³⁷⁵, no phosphorylation of Thr³⁷⁶ or Thr³⁷⁹ (Just et al., 2013) and less arrestin translocation (McPherson et al., 2010). In the present study we observed that Cmpd101 partially inhibited morphine-induced MOPr desensitization in LC neurons.

Our studies with Cmpd101 support the view that GRK2 and GRK3 do play a role in MOPr desensitization in LC neurons. What was surprising is that when we calculated the loss of MOPr function resulting from agonist-induced desensitization we found that Cmpd101 only partially reversed the loss of MOPr function that underlies the desensitization. The inability of Cmpd101 to fully reverse the loss of MOPr function induced by DAMGO could indicate either that there are two mechanisms of MOPr desensitization – one involving GRK mediated phosphorylation (inhibited by Cmpd101) and one that does not involve GRK2 and GRK3 – or that, in intact neurons, for reasons that are unclear, Cmpd101 does not completely inhibit GRK2 and GRK3. Evidence against the incomplete GRK inhibition hypothesis would be that in HEK 293 cells Cmpd101 very effectively reduced DAMGO-induced MOPr phosphorylation, arrestin translocation and MOPr internalization. However, one caveat to this is that, by necessity the phosphorylation, translocation and internalization studies were performed under very different experimental conditions to the electrophysiological experiments. If there is a second mechanism involved in MOPr desensitization this is

currently unidentified, but it does not involve ERK1/2, PKC or JNK as in the present study inhibitors of these kinases did not reduce MOPr desensitization further in the presence of Cmpd101. One potential alternative mechanism for the residual component of MOPr desensitization is that catalytically-inactive GRK2/3 binds $\beta\gamma$ subunits of the G-protein, inhibiting GIRK channel function (Raveh et al., 2010), while we cannot rule this out previous work suggests that this process does not play a role in rat LC neurons of this age (Llorente et al., 2012). Of the other GRK subtypes expressed in mammalian neurons (GRK5 and GRK6), our data suggest that GRK5 is not responsible, as the GRK5 inhibitor, amlexanox, was ineffective. Although amlexanox lacks specificity (Reilly et al., 2013), and we have no direct proof that it is permeating LC neurons in the slice, it has been shown to inhibit intracellular GRK5-mediated responses, with near complete block at 50 μ M (Homan et al., 2014b). It is not currently known if amlexanox also inhibits the structurally-similar GRK6, although previous studies (Bailey et al., 2009b) have shown no effect of GRK6 on MOPr desensitization using viral-transfection of dominant negative mutant GRK6.

We have been unable to obtain any evidence to support the view that ERK1/2 participate in MOPr desensitization as proposed by Dang et al., (2009). In our preliminary *in vitro* kinase screen Cmpd101 did show some inhibition of both ERK1 and ERK2 but this was modest (20% inhibition) with a concentration of Cmpd101 that would inhibit GRK2 by over 90%. However we found no evidence in intact cells that Cmpd101 could decrease ERK1/2 phosphorylation or inhibit the phosphorylation of Elk-1 by ERK1/2. Thus we have excluded the possibility that Cmpd101 inhibits MOPr desensitization by inhibiting both GRK2/3 and ERK1/2.

In conclusion, we demonstrate that Cmpd101 offers a simple means to study the roles of GRK2/3 in GPCR desensitization and other cellular functions. We show here that a component of MOPr desensitization in rodent LC neurons is GRK2/3-mediated. Data from GRK3 knockout mice demonstrate that GRK3 ablation alone is insufficient to inhibit MOPr

desensitization in these neurons, suggesting either a selective role for GRK2, or, redundancy of action between GRK2 and GRK3. The development of a membrane-permeable, small-molecule inhibitor of GRK2/3 gives the opportunity to explore the roles of these kinases in desensitization of MOPRs, and other GPCRs, in native tissues and *in vivo*.

Acknowledgements

The authors thank John Tesmer (University of Michigan) for help and guidance with the synthesis of Compound101.

Authorship contributions

Participated in research design: *Bailey CP, Kelly E, Lowe JD and Henderson G.*

Conducted experiments: *Cooke AE, Kelly E, Lowe JD, Sanderson HS, Withey SL and Tsisanova E*

Contributed new reagents or analytic tools: *Chavkin C, Husbands SM and Ostovar M*

Performed data analysis: *Cooke AE, Kelly E, Lowe JD, Sanderson HS and Tsisanova E.*

Wrote or contributed to the writing of the manuscript: *Lowe JD, Kelly E, Henderson G and Bailey CP.*

References

Alvarez VA, Arttamangkul S, Dang V, Salem A, Whistler JL, Von Zastrow M, Grandy DK, Williams JT (2002) μ -Opioid receptors: Ligand-dependent activation of potassium conductance, desensitization, and internalization. *J Neurosci* **22**:5769-5776.

Arttamangkul S, Quillinan N, Low MJ, von Zastrow M, Pintar J, Williams JT (2008) Differential activation and trafficking of micro-opioid receptors in brain slices. *Mol Pharmacol* **74**:972-979.

Arttamangkul S, Lau EK, Lu HW, Williams JT (2012) Desensitization and trafficking of μ -opioid receptors in locus ceruleus neurons: modulation by kinases. *Mol Pharmacol* **81**:348-355.

Arttamangkul S, Birdsong W, Williams JT (2015) Does PKC activation increase the homologous desensitization of μ opioid receptors? *Br J Pharmacol* **172**:583-592.

Bailey CP, Couch D, Johnson E, Griffiths K, Kelly E, Henderson G. (2003) μ -opioid receptor desensitization in mature rat neurons: lack of interaction between DAMGO and morphine. *J Neurosci* **23**:10515-10520.

Bailey CP, Kelly E, Henderson G (2004) Protein kinase C activation enhances morphine-induced rapid desensitization of μ -opioid receptors in mature rat locus ceruleus neurons. *Mol Pharmacol* **66**:1592-1598.

Bailey CP, Oldfield S, Llorente J, Caunt CJ, Teschemacher AG, Roberts L, McArdle CA, Smith FL, Dewey WL, Kelly E, Henderson G (2009a) Involvement of PKC α and G protein-coupled receptor kinase 2 in agonist-selective desensitization of μ -opioid receptors in mature brain neurons. *Br J Pharmacol* **158**:157-164.

Bailey CP, Llorente J, Gabra BH, Smith FL, Dewey WL, Kelly E, Henderson G (2009b) Role of protein kinase C and mu-opioid receptor (MOPr) desensitization in tolerance to morphine in rat locus coeruleus neurons. *Eur J Neurosci* **29**:307-318.

Bain J, Plater L, Elliott M, Shpiro N, Hastie CJ, McLauchlan H, Klevernic I, Arthur JS, Alessi DR, Cohen P (2007) The selectivity of protein kinase inhibitors: a further update. *Biochem J* **408**:297-315.

Black JW, Leff P, Shankley NP, Wood J (1985) An operational model of pharmacological agonism: the effect of E/[A] curve shape on agonist dissociation constant estimation. *Br J Pharmacol* **84**:561-571.

Chen YJ, Oldfield S, Butcher AJ, Tobin AB, Saxena K, Gurevich VV, Benovic JL, Henderson G, Kelly E (2013) Identification of phosphorylation sites in the COOH-terminal tail of the μ -opioid receptor. *J Neurochem* **124**:189-199.

Connor M, Osborne PB, Christie MJ (2004) Mu-opioid receptor desensitization: is morphine different? *Br J Pharmacol* **143**:685-696.

Cruzalegui FH, Cano E, Treisman R (1999) ERK activation induces phosphorylation of Elk-1 at multiple S/T-P motifs to high stoichiometry. *Oncogene* **18**:7948-7957.

Dang VC, Napier IA, Christie MJ (2009) Two distinct mechanisms mediate acute mu-opioid receptor desensitization in native neurons. *J Neurosci* **29**:3322-3327.

Doll C, Konietzko J, Pöll F, Koch T, Höllt V, Schulz S (2011) Agonist-selective patterns of μ -opioid receptor phosphorylation revealed by phosphosite-specific antibodies. *Br J Pharmacol* **164**:298-307.

Doll C, Pöll F, Peuker K, Loktev A, Glück L, Schulz S (2012) Deciphering μ -opioid receptor phosphorylation and dephosphorylation in HEK 293 cells. *Br J Pharmacol* **167**:1259-1270.

Erdtmann-Vourliotis M, Mayer P, Ammon S, Riechert U, Höllt V (2001) Distribution of G protein-coupled receptor kinase (GRK) isoforms 2, 3, 5 and 6 mRNA in the rat brain. *Brain Res Mol Brain Res* **95**:129-137.

Groer CE, Schmid CL, Jaeger AM, Bohn LM (2011) Agonist-directed interactions with specific beta-arrestins determine mu-opioid receptor trafficking, ubiquitination, and dephosphorylation. *J Biol Chem* **286**:31731-31741.

Hastie CJ, McLauchlan HJ, Cohen P (2006) Assay of protein kinases using radiolabeled ATP: a protocol. *Nat Protoc* **1**:968-971.

Homan KT, Wu E, Wilson MW, Singh P, Larsen SD, Tesmer JJ (2014a) Structural and functional analysis of G protein-coupled receptor kinase inhibition by paroxetine and a rationally designed analog. *Mol Pharmacol* **85**:237-248.

Homan KT, Wu E, Cannavo A, Koch WJ, Tesmer JJ (2014b) Identification and characterization of amlexanox as a G protein-coupled receptor kinase 5 inhibitor. *Molecules* **19**:16937-16949.

Hull LC, Llorente J, Gabra BH, Smith FL, Kelly E, Bailey C, Henderson G, Dewey WL (2010) The effect of protein kinase C and G protein-coupled receptor kinase inhibition on tolerance induced by mu-opioid agonists of different efficacy. *J Pharmacol Exp Ther* **332**:1127-1135.

Iino M, Furugori T, Mori T, Moriyama S, Fukuzawa A, Shibano T (2002) Rational design and evaluation of new lead compound structures for selective betaARK1 inhibitors. *J Med Chem* **45**:2150-2159.

Ikeda S, Keneko M, Fujiwara S (2007), inventors; Takeda Pharmaceutical Company Ltd., Ikeda S, Keneko M, Fujiwara S, assignees. Cardiotonic agent comprising GRK inhibitor. World patent WO2007034846. 2007 March 29.

Johnson EA, Oldfield S, Braksator E, Gonzalez-Cuello A, Couch D, Hall KJ, Mundell SJ, Bailey CP, Kelly E, Henderson G (2006) Agonist-selective mechanisms of mu-opioid receptor desensitization in human embryonic kidney 293 cells. *Mol Pharmacol* **70**:676-685.

Kelly E, Bailey CP, Henderson G (2008) Agonist-selective mechanisms of GPCR desensitization. *Br J Pharmacol* **153 Suppl 1**:S379-388.

Just S, Illing S, Trester-Zedlitz M, Lau EK, Kotowski SJ, Miess E, Mann A, Doll C, Trinidad JC, Burlingame AL, von Zastrow M, Schulz S (2013) Differentiation of opioid drug effects by hierarchical multi-site phosphorylation. *Mol Pharmacol* **83**:633-639.

Li AH, Wang HL (2001) G protein-coupled receptor kinase 2 mediates mu-opioid receptor desensitization in GABAergic neurons of the nucleus raphe magnus. *J Neurochem* **77**:435-444.

Llorente J, Lowe JD, Sanderson HS, Tsisanova E, Kelly E, Henderson G, Bailey CP (2012) μ -Opioid receptor desensitization: homologous or heterologous? *Eur J Neurosci* **36**:3636-3642.

Lohse MJ, Benovic JL, Caron MG, Lefkowitz RJ (1990) Multiple pathways of rapid beta 2-adrenergic receptor desensitisation. Delineation with specific inhibitors. *J Biol Chem* **265**:3202-3211.

Manning G, Whyte DB, Martinez R, Hunter T, Sudarsanam S (2002) The protein kinase complement of the human genome. *Science* **298**:1912-1934.

McPherson J, Rivero G, Baptist M, Llorente J, Al-Sabah S, Krasel C, Dewey WL, Bailey CP, Rosethorne EM, Charlton SJ, Henderson G, Kelly E (2010) μ -opioid receptors: correlation of agonist efficacy for signalling with ability to activate internalization. *Mol Pharmacol* **78**:756-766.

- Mittal N, Tan M, Egbuta O, Desai N, Crawford C, Xie CW, Evans C, Walwyn W (2012) Evidence that behavioral phenotypes of morphine in β -arr2^{-/-} mice are due to the unmasking of JNK signaling. *Neuropsychopharmacology* **37**:1953-1962.
- Mundell SJ, Luo J, Benovic JL, Conley PB, Poole AW (2006) Distinct clathrin-coated pits sort different G protein-coupled receptor cargo. *Traffic* **7**:1420-1431.
- North RA, Williams JT (1985) On the potassium conductance increased by opioids in rat locus coeruleus neurones. *J Physiol* **364**:265-280.
- Quillinan N, Lau EK, Virk M, von Zastrow M, Williams JT (2011) Recovery from mu-opioid receptor desensitization after chronic treatment with morphine and methadone. *J Neurosci* **31**:4434-4443.
- Raveh A, Cooper A, Guy-David L, Reuveny E (2010) Nonenzymatic rapid control of GIRK channel function by a G protein-coupled receptor kinase. *Cell*, **143**:750–760.
- Reilly SM, Chiang SH, Decker SJ, Chang L, Uhm M, Larsen MJ, Rubin JR, Mowers J, White NM, Hochberg I, Downes M, Yu RT, Liddle C, Evans RM, Oh D, Li P, Olefsky JM, Saltiel AR (2013) An inhibitor of the protein kinases TBK1 and IKK- ϵ improves obesity-related metabolic dysfunctions in mice. *Nat Med* **19**:313-321.
- Roskoski R (2012) ERK1/2 MAP kinases: Structure, function, and regulation. *Pharm Res* **66**: 105-143.
- Shenoy SK, Lefkowitz RJ (2011) β -Arrestin-mediated receptor trafficking and signal transduction. *Trends Pharmacol Sci* **32**:521-533.
- Terman GW, Jin W, Cheong YP, Lowe J, Caron MG, Lefkowitz RJ, Chavkin C (2004) G protein receptor kinase 3 (GRK3) influences opioid analgesic tolerance but not opioid withdrawal. *Br J Pharmacol* **141**:55-64.

Thal DM, Yeow RY, Schoenau C, Huber J, Tesmer JJ (2011) Molecular mechanism of selectivity among G protein-coupled receptor kinase 2 inhibitors. *Mol Pharmacol* **80**:294-303.

Thal DM, Homan KT, Chen J, Wu EK, Hinkle PM, Huang ZM, Chuprun JK, Song J, Gao E, Cheung JY, Sklar LA, Koch WJ, Tesmer JJ (2012) Paroxetine is a direct inhibitor of G protein-coupled receptor kinase 2 and increases myocardial contractility. *ACS Chem Biol* **7**:1830-1839

Walwyn W, Evans CJ, Hales TG (2007) Beta-arrestin2 and c-Src regulate the constitutive activity and recycling of mu opioid receptors in dorsal root ganglion neurons. *J Neurosci* **27**:5092-5104.

Williams JT, Ingram SL, Henderson G, Chavkin C, von Zastrow M, Schulz S, Koch T, Evans CJ, Christie MJ (2013) Regulation of μ -opioid receptors: desensitization, phosphorylation, internalization, and tolerance. *Pharmacol Rev* **65**:223-54.

Footnotes

Financial support.

Aspects of this work were supported by grants from the Medical Research Council, UK [MR/J013269/1] and the Biotechnology and Biological Sciences Research Council, UK [BB/J003506/1] to E.P.K. and G.H., from the Wellcome Trust, UK [Value in People award] to J.D.L. and C.P.B., and from the United States Public Health Service – National Institute on Drug Abuse [RO1DA030074] to C.C.

Figure Legends

Figure 1. Inhibition of MOPr desensitization by Compound 101 in rat LC neurons.

Traces **A, D, G and J** show outward potassium currents recorded from rat LC neurons in response to a receptor saturating concentrations of methionine enkephalin (Met Enk, 30 μ M), DAMGO (10 μ M), endomorphin-2 (10 μ M) and morphine (morph, 30 μ M). The Met Enk, DAMGO and endomorphin-2 responses desensitized rapidly over the 10 min of agonist application. The desensitization induced by morphine was less than that produced by the other agonists and was measured after 15 min of morphine application. Agonist responses returned to baseline after wash out (Met Enk) or when naloxone (nalox, 1 μ M) was applied. The middle column of traces (**B, E, H, K**) show currents induced by each agonist in slices exposed to Compound 101 (Cmpd101, 30 μ M) for at least 15 min before and during the application of the opioid agonists. **C, F, I and L** show pooled data for the percentage desensitization after 10 min of Met Enk, DAMGO or endomorphin-2 application and 15 min of morphine application from experiments such as those illustrated in the two columns of experimental traces. Cmpd101 significantly inhibited the desensitization of all four agonists (Met Enk: n=5 for all experiments; DAMGO: n=4 for all experiments; endomorphin-2: n=6 for all experiments; morphine: n=6 for all experiments) In all histograms * indicates $p < 0.05$, ANOVA compared to the appropriate control). All scale bars 50 pA, 2 min.

Figure 2. Effect of GRK3 knockout and Compound 101 on methionine enkephalin-induced MOPr desensitization in mouse LC neurons.

A, Representative potassium currents in response to a receptor saturating concentration of Methionine enkephalin (Met Enk, 30 μ M) recorded from mouse LC neurons in slices taken from either wildtype (WT, upper) or GRK3 knockout (KO, lower) mouse brains. **B**, Pooled data from experiments as illustrated in A. The desensitization induced by Met Enk over a 10 min application was not different in GRK3 KO mice compared to WT littermate controls (GRK3 WT, n=6; GRK3 KO, n=6, t-test, $p > 0.05$). **C**, Representative potassium currents in response to Met Enk (30 μ M)

recorded from LC neurons from slices taken from C57Bl/6J mice that were either untreated (upper) or pre-treated with Cmpd101 (30 μ M) for at least 15 min prior to and during the application of Met Enk (lower). **D**, Pooled data for the percentage desensitization over the 10 min of opioid agonist application from experiments such as those illustrated in C. Cmpd101 (3 and 30 μ M) significantly inhibited Met Enk-induced desensitization measured after 10 min of agonist application. (n=5 for each; * indicates $p<0.05$, ANOVA compared to control). All scale bars 15 pA, 2 min.

Figure 3. Compound 101 reduced the depression of the maximum response to morphine produced by DAMGO-induced desensitization. **A & B**, Representative potassium current traces showing the amplitude of the maximum response to morphine (morph) compared to that of noradrenaline (NA, 100 μ M) in the absence (**A**) or after induction of desensitization (**B**) induced by application of DAMGO (10 μ M for 12 min). The opioid antagonist naloxone (nalox, 1 μ M) was added after morphine to bring the response back to baseline prior to application of NA. **C**, Representative current trace from an experiment in which slices were exposed to Compound 101 (Cmpd101, 30 μ M) for at least 15 min before and during the application of the opioid agonists. **D**, Pooled data from experiments as illustrated in **A**, **B** & **C**. DAMGO-induced desensitization inhibited the maximum response to morphine. Cmpd101 concentration-dependently reversed the DAMGO-desensitization-induced decrease in the morphine response (n=4 for all experiments; * indicates $p<0.05$, ANOVA). All scale bars 60 pA, 2 min.

Figure 4. Inhibition of DAMGO-induced MOPr phosphorylation and arrestin recruitment by compound 101. **A**, HEK 293 cells stably expressing HA-tagged rat MOPr were pre-treated with Cmpd101 for 30 min prior to stimulation with DAMGO (10 μ M for 5 min). Agonist-induced phosphorylation was assessed by Western blot analysis using an

antibody targeting phospho-Ser³⁷⁵ (pS375). Anti-HA and anti-tubulin antibodies confirmed equal loading of the gels. **B**, Western blots as illustrated in **A** were quantified by densitometry and expressed as a percentage of the maximal phosphorylation in response to DAMGO (10 μ M) in each experiment. Cmpd101 (30 μ M) abolished DAMGO-induced MOPr phosphorylation at Ser³⁷⁵ (n=3; * indicates $p<0.05$, ANOVA compared to Control + DAMGO). No phosphorylation was seen under control conditions or with Cmpd101 alone (n=3). **C**, DAMGO-induced arrestin3 translocation to the receptor was measured using the DiscoverX PathHunter assay. DAMGO (10 μ M) application produced a robust recruitment of arrestin3 to the receptor that was significantly inhibited in cells that were pre-treated with Cmpd101 (30 μ M) for 30 min (n=3, * indicates $p<0.05$, ANOVA). **D**, Internalization of HA-MOPrs expressed in HEK 293 cells assessed by ELISA using an anti-HA antibody to label surface receptors. DAMGO (10 μ M) induced a time-dependent loss of surface receptors that was significantly inhibited by Cmpd101 (n=3-4, * indicates $p<0.05$, ANOVA compared to control). **E**, Confocal images of HA-MOPrs following incubation with anti-HA antibody and fluorescein-tagged secondary antibody (green), counterstained with Hoechst 33258 nucleic acid stain (blue) following incubation with DAMGO (10 μ M) and/or Cmpd101 (30 μ M). Images are from one experiment repeated 3 times. Scale bar = 10 μ M.

Figure 5. Effect of Compound 101 on the phosphorylation of ERK1/2 and Elk-1. **A**, ERK1/2 activity in HEK 293 cells expressing HA-MOPrs was assessed by Western blot using an antibody targeting phospho-ERK1/2. DAMGO (10 μ M) application for 5 min produced a robust phosphorylation of ERK1/2 both in control cells and cells that had been pre-treated with Cmpd101 for 30 min. Antibodies against total ERK1/2 and tubulin confirmed equal loading of the gels. **B**, Western blots as illustrated in **A** were quantified by densitometry and expressed as a percentage of the maximal ERK1/2 phosphorylation in response to DAMGO (10 μ M) in each experiment. Cmpd101 did not affect DAMGO-induced ERK1/2 phosphorylation ($p>0.05$, ANOVA), but at the higher 30 μ M concentration, Cmpd101

produced a modest but significant activation of ERK1/2 on its own ($p < 0.05$, one-sample t-test versus control). $n=4$ for each. **C**, Phosphorylation of the ERK1/2 substrate Elk-1 in response to DAMGO was assessed by Western blot using an antibody targeting pSer383 [of Elk-1]. Treatment of HA-MOPr cells with DAMGO ($10 \mu\text{M}$) for 5 min produced phosphorylation of Elk-1 which was unaffected by pre-treatment with Cmpd101 ($30 \mu\text{M}$) for 30 min. In contrast, agonist dependent phosphorylation of both Elk-1 and ERK1/2 was significantly reduced following pre-treatment for 30 min with the MEK1 inhibitor PD98059 ($10 \mu\text{M}$). Antibodies against total ERK1/2 and tubulin confirmed equal loading of the gels. **D**, Western blots of pELK-1 as shown in C were quantified by densitometry and expressed as a percentage of the maximal Elk-1 phosphorylation in response to DAMGO ($10 \mu\text{M}$) in each experiment. Cmpd101 did not reduce DAMGO-induced Elk-1 phosphorylation at a concentration of $30 \mu\text{M}$. $n=3$. **E**, The ERK1/2 inhibitor, PD98085 ($10 \mu\text{M}$), reduced both DAMGO-induced pELK-1 and pERK1/2 activation to basal levels. Data as in C quantified by densitometry. * signifies $p < 0.05$, one-sample t-test versus control. $n=3$.

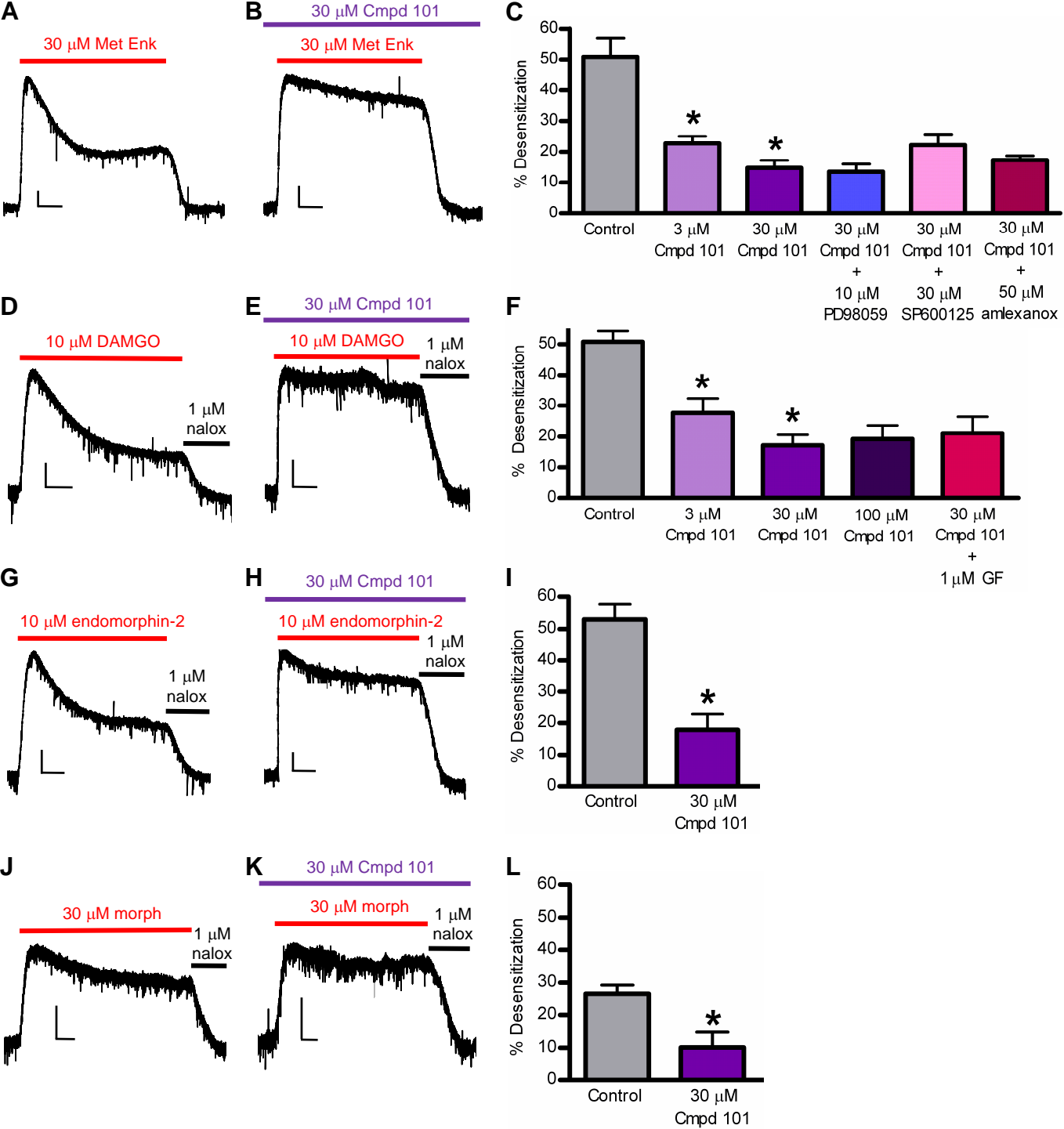


Figure 1

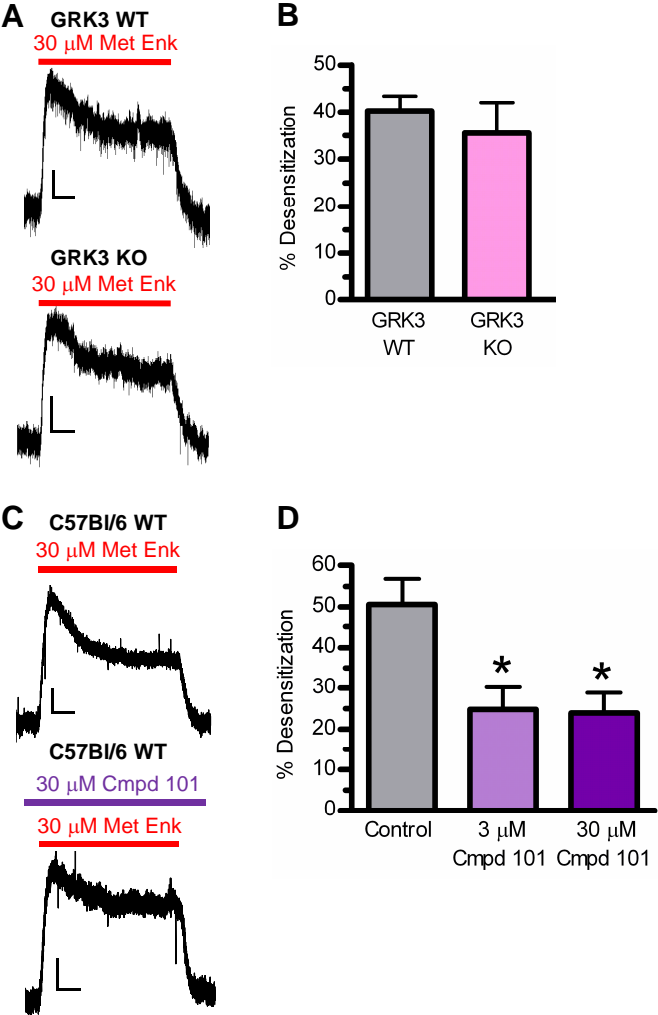


Figure 2

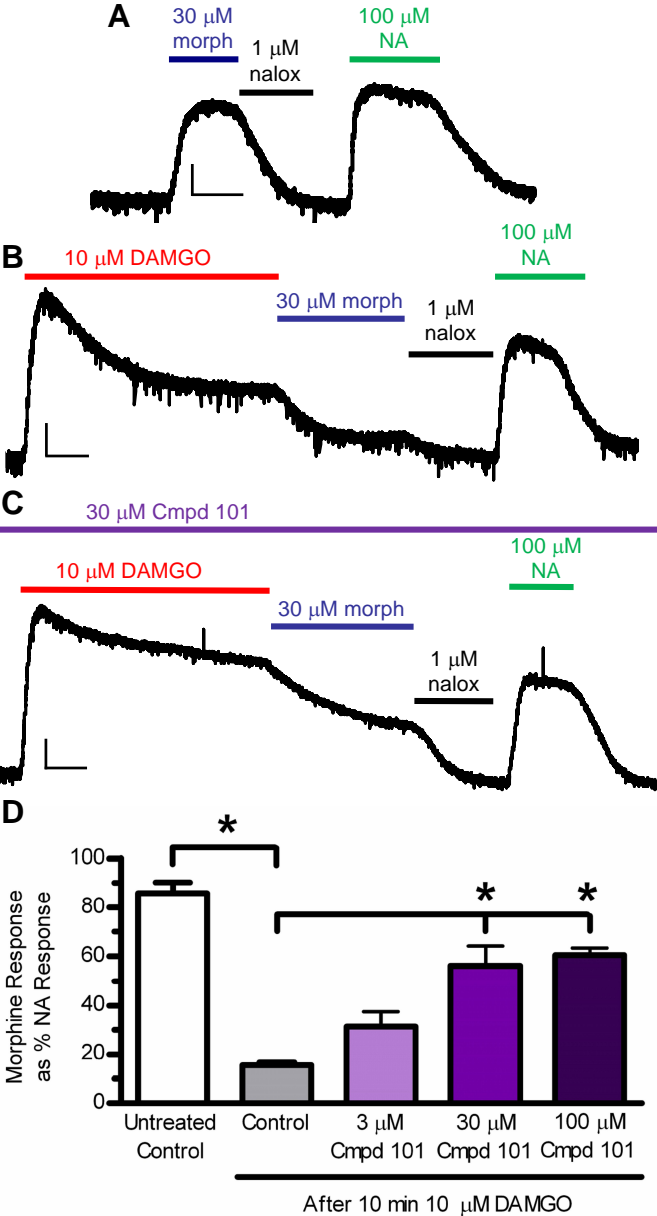


Figure 3

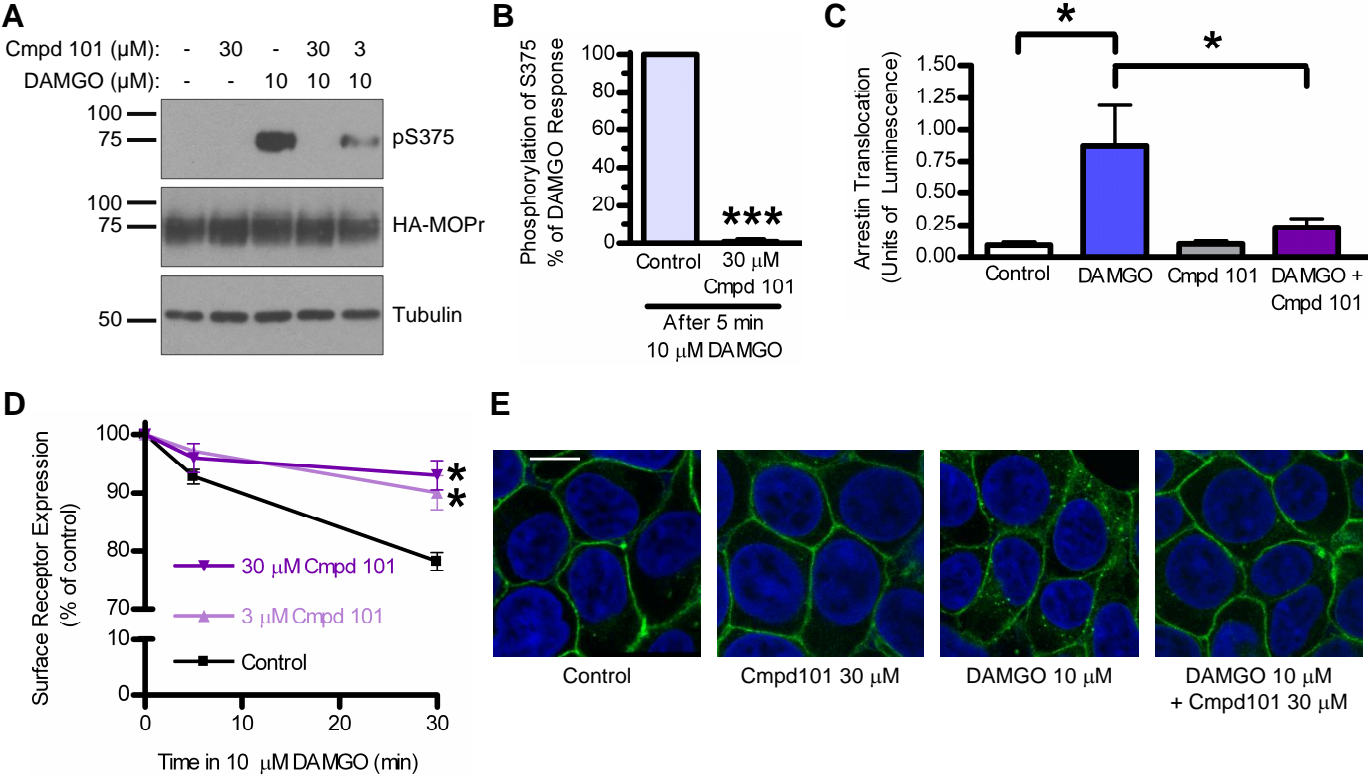


Figure 4

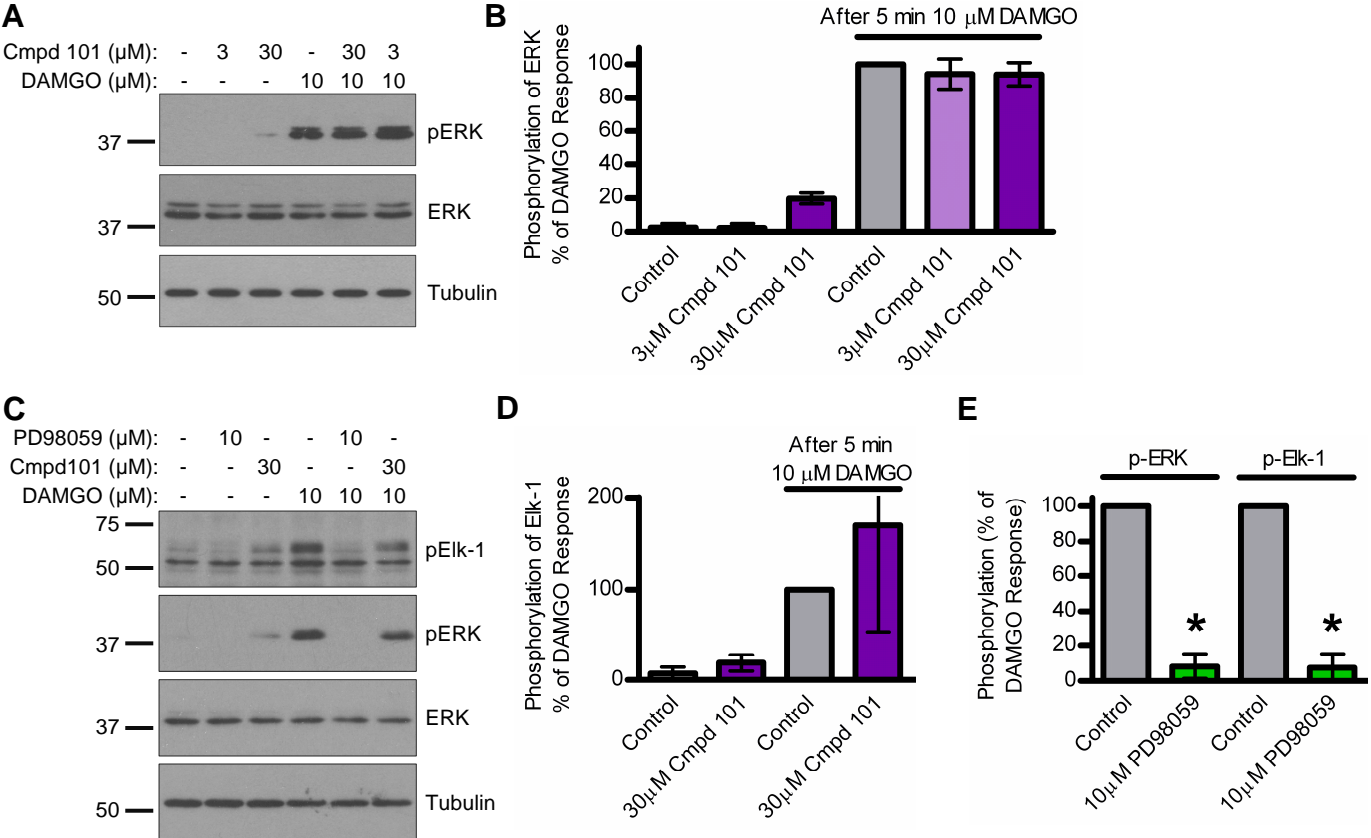


Figure 5

Supplementary Data

**Role of G protein-coupled receptor kinases 2 and 3 in μ -opioid receptor
desensitization and internalization**

by

**Janet D. Lowe, Helen S. Sanderson, Alexandra E. Cooke, Mehrnoosh Ostovar,
Elena Tsisanova, Sarah L. Withey, Charles Chavkin, Stephen M. Husbands,
Eamonn Kelly, Graeme Henderson and Chris P. Bailey**

School of Physiology and Pharmacology, University of Bristol, Bristol, United Kingdom
(J.D.L., H.S.S., A.E.C., E.T., S. L. W., E.K., G.H.); Department of Pharmacology,
University of Washington School of Medicine, Seattle, WA, USA (C.C.); Department of
Pharmacy and Pharmacology, University of Bath, Bath, United Kingdom (M.O., S.H.,
C.P.B.)

MOLECULAR PHARMACOLOGY

Supplementary Data Table 1.**Inhibition of protein kinases by Compound 101**

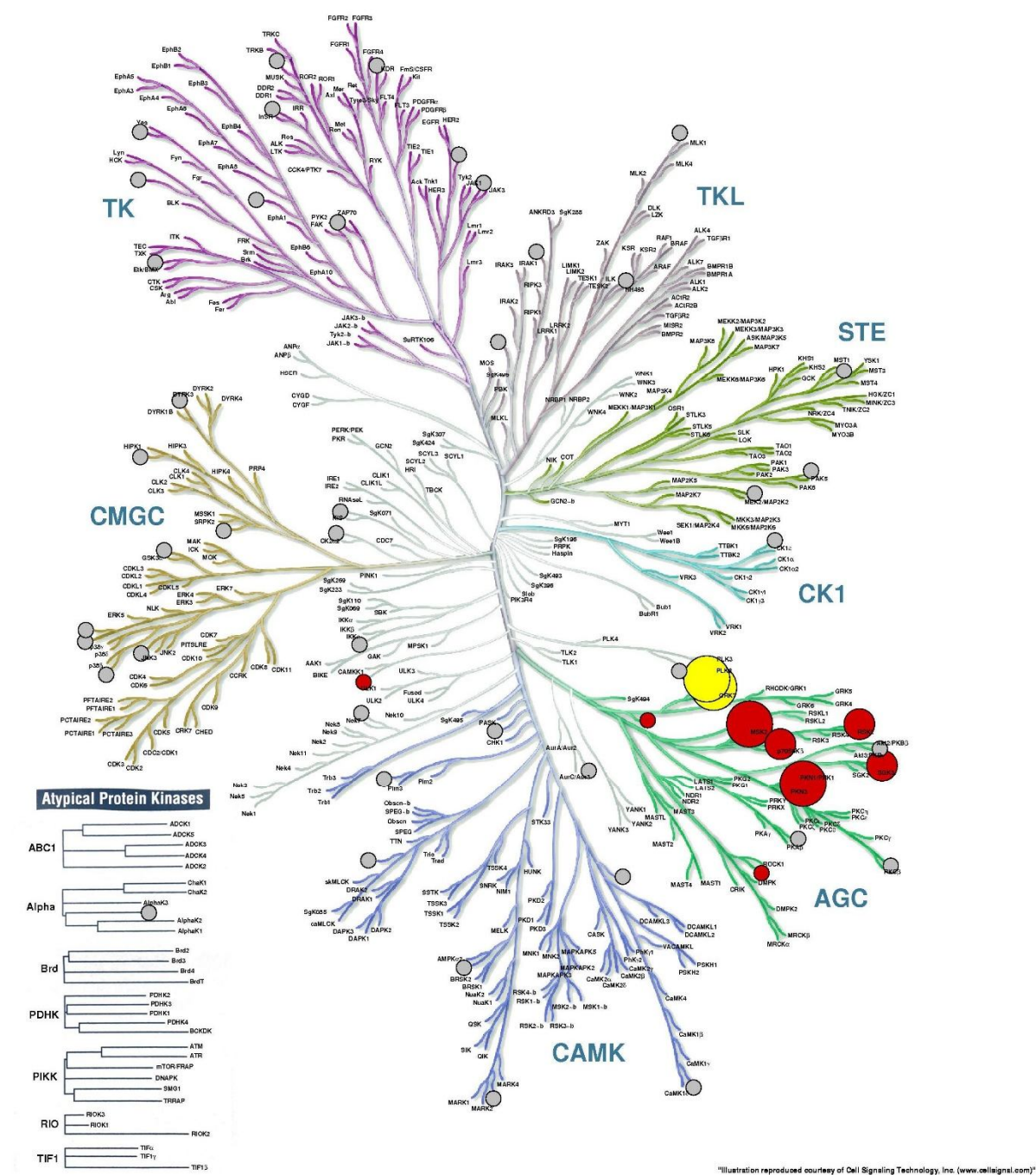
Cmpd101 was studied at a concentration of 1 μ M. Results are given as the percentage of kinase activity in the absence of the inhibitor. The screen was performed in duplicate and results are given as a mean value with standard deviation.

Kinase	% Activity Remaining	St Dev
PRK2	7	1
MSK1	10	1
SGK1	31	5
S6K1	47	4
RSK1	48	5
ROCK 2	53	1
PKD1	65	1
CAMKKb	69	3
BTK	72	1
VEG-FR	79	5
CHK2	79	4
ERK2	81	3
AMPK (hum)	82	14
PKBa	83	3
ERK1	83	2
TBK1	84	7
EPH-A2	85	6
GSK3b	85	6
MLK3	86	4
HER4	86	23
CK1 δ	86	8
MKK1	88	11
MST2	89	8
SmMLCK	89	5
TTK	89	3
LKB1	91	5
SRPK1	91	11
Aurora B	92	3
MARK3	92	7
RIPK2	93	10
PDK1	93	5
JNK1	93	5
Lck	94	2
JAK2	94	3
PKA	95	2
NEK6	95	6
PKCa	96	3
IRAK4	96	4
CAMK1	101	13

MOL #98293

PAK4	101	8
p38a MAPK	101	7
SYK	103	8
IGF-1R	103	3
HIPK2	103	13
EF2K	103	15
Src	104	12
TrkA	106	7
CK2	106	6
PLK1	106	2
PIM1	107	7
DYRK1A	112	3
TAK1	113	11

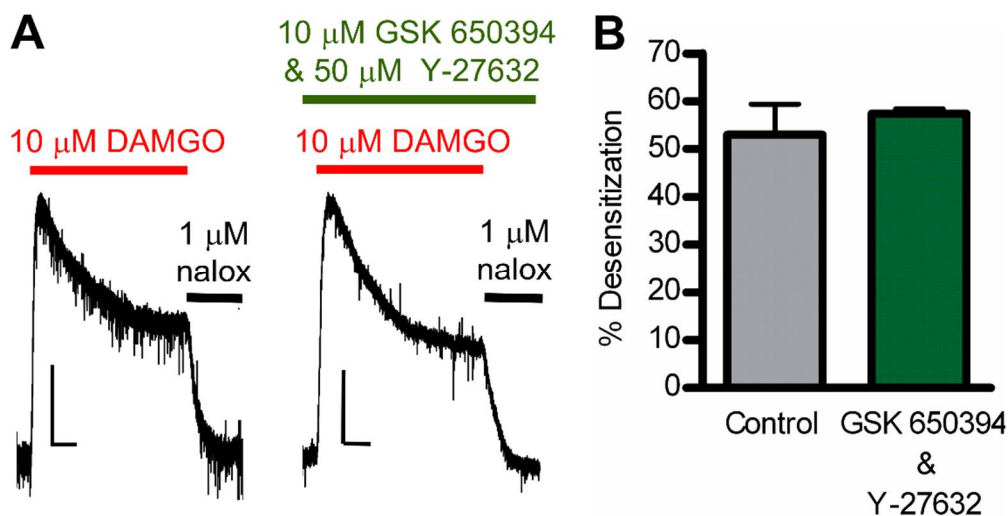
Supplementary Data Figure 1.



Kinome tree highlighting the primary targets of Cmpd101. Phylogenetic representation of the human kinome is annotated showing the primary targets of Cmpd 101 as GRK2/3 (yellow circles) (Manning et al., 2002). The additional kinases we identified that are inhibited by Cmpd 101 *in vitro* (1 μ M) are also highlighted (red

circles). Diameter is proportional to the activity remaining; large red circle 0-25%, medium 25-50% and small 50-70%. Grey circles indicate kinases that were tested in the screen but whose remaining activity was >70%. The tree indicates broad sampling from across the kinome. The tree was generated using Kinome Render and the illustration reproduced courtesy of Cell Signaling Technology, Inc. (www.cellsignal.com).

Supplementary Data Figure 2.



DAMGO-induced MOPr desensitization was unaffected by inhibition of off-target kinases inhibited by Compound 101.

A, Representative potassium current traces from either untreated rat LC neurons or neurons pre-treated with GSK650394 (10 μ M) and Y-27632 (50 μ M) for at least 15 min prior to and during application of DAMGO (10 μ M).

B, Pooled data from experiments as illustrated in **A**. The combination of kinase inhibitors which inhibit SGK1 (GSK650394), PRK2 and ROCK2 (Y-27632) did not affect DAMGO-induced desensitization (n=4 for both, p>0.05, t-test,). All scale bars 60 pA, 2 min.